1,6 hyd (FbrfOO75214)

same as Cheney, Hopkins 2yr 328 New record

**DROSOPHILA DATABASE REPORT FORM**
Return to John Merriam, UCLA Biology, LA 90024-1606 FAX [redacted]

Please verify [✓] or correct the address and citation on the reverse.

**NAME:** Liz Mansfield PHONE #: (301) 338-7717 FAX #: [redacted]

1. **GENE:** How do you describe the gene you are reporting on?

   mutant name (if any): hyperplastic discs

   abbreviation: hyd

   biochemical name (if any): _____

2. **LOCATION:** Where is your clone/walk located? _____
   (If you are using a clone from a more extended walk, what is the location of your gene? 85E )

3. **SIZE:** How many kb did you clone? (round to nearest kb)

   genomic 30 vector/library name: Tamkun cosmid library (CosPer vector)

   cDNA _____ vector/library name: _____

4. **METHOD:** Describe the entry point for your clone/walk.

   prior clone from α-tubulin transposon tag X P mutants? yes, 3 insertions

   expression _____ differential screen _____

   microclone _____ oligonucleotide _____ PCR? _____

   homology _____

5. **SEQUENCE:** How many kb did you sequence? (round to nearest 0.1 kb)

   genomic _____ accession #: _____

   cDNA _____ accession #: _____

   PRODUCT: How many amino acids in the ORF? _____

   (List above the predicted function or homology group of the ORF)

   (List above the key words that describe domains in the ORF)

6. **TRANSCRIPTS:** (round mRNA sizes to nearest 0.1 kb, list largest 1st):
   ≥12 Kb, beyond MW marker range When/where found? 6,7 day larvae polyA+ & total

7. **PROTEIN** (antibody detection): Where found? _____

8. **LAC Z FUSION** stain: Where found? _____

9. **MORE DETAILS:** l(3)c43

Arthur Shaver, Barbara

1990 Crete 328

**DROSOPHILA DATABASE REPORT FORM**
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Please verify [___] or correct the address and citation on the reverse.

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   expression_____ differential screen_____

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   (List above the key words that describe domains in the ORF)

6. **TRANSCRIPTS:** (round mRNA sizes to nearest 0.1 kb, list largest 1st):

   _____ When/where found?_____

7. **PROTEIN** (antibody detection): Where found?_____

8. **LAC Z FUSION** stain: Where found?_____

9. **MORE DETAILS:** proximal to 85 alpha tubulin

   1500, 1055, 910, 820, 609, 328, 327, 209, 74

D

dc

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UCLA

328

SANTA BARBARA • SANTA CRUZ

December 20, 1990

FAX: [redacted]

Mansfield, Liz Shearn
Biology Department
Johns Hopkins University
34th and Charles Streets
Baltimore, MD 21218

John Merriam
UCLA Drosophila Genmaps Database
DEPARTMENT OF BIOLOGY
405 HILGARD AVENUE
LOS ANGELES, CALIFORNIA 90024-1606
FAX [redacted]

Dear Colleagues:

I would like to use information from the clone list to construct a "physical map" of the Drosophila genome. You have (a) clone(s) listed, as described below, so I would like to enlist your help. What I need for this project are the names and locations of chromosome rearrangement breakpoints that lie within the region of your clone/walk. Mapping breakpoints is the most reliable way of connecting the molecular with the cytological scales.

As you know, the clone list has been useful to Drosophila workers in finding starting points for their walks. The database is [illegible] references, so that the original cloning paper from a lab is cited as key #1, with subsequent papers cited by subject (key #5 is sequence information, for example). The clone list is therefore also a gene-by-gene literature index. The clone list is now mounted on the Indiana University BioArchive computer bulletin board in the FLY subdirectory. Use Internet address 129.79.1.101 and your local File Transfer Program.

Please take a few minutes to complete this form and return to me by FAX or mail. It will be most useful if you can update your clone sizes and make more exact your clone locations. Make any other changes you want also. List the rearrangements and which breakpoints you have identified. Any cytologically visible rearrangement (deletion, inversion, translocation, transposition, duplication) is useful. Please indicate if the same citation is the best reference, or indicate a better one (or a personal communication).

I thank you for your help as always. With your prompt and generous support I can expect to show you the results in March at the Chicago fly meeting.

[redacted]

| List Rearrangements: | Breakpoint found: | Other breakpoints: |
|---|---|---|
| Df (3R) hyd $^{XB104}$ | 85E5 | |
| Df (3R) by $^{446}$ | 85E1 | |
| | | |

CHANGE or CONFIRM indexing for original clone cit'n: Locations X Clone sizes __ Names/Products __

Mansfield, L; Biggs, J; Shearn, A. 1990 Crete meeting.

| Location | Key | kb gen/cDNA | Gene name (abbreviation) product rearrangements... |
|---|---|---|---|
| 85E | 1,6 | 30 0 | hyperplastic discs (hyd) |

Location is now narrowed to 85E1-5 on the basis of deletion complementation: )

hyd

source_number: 328

full_location: 85E  cross_locations:

table_numbers: 1,6

gene_name: hyperplastic discs

gene_abbrev: hyd

product:  YACs: 0

kb_cloned_genom: 30  vector_genomic: CosPer (Tankun cosmid library)

kb_cloned_cDNA: 0  vector_cDNA:

expression:

homology:

microcloning:

transposon_tag: P mutants -- 3 insertions

oligonucleotide:

clonetechtext:

rearrangement:

T_strain:

markers:

kb_seq_genomic: 0  acc_number_gen:  kb_seq_old: 0

kb_seq_cDNA: 0  acc_number_cDNA:

comments:

ORF_size:

ORF_function:

ORF_domains:

kb_trans_size: 12

transc_dist: 6-7 day larvae polyA+ and total

protein_dist:

fusion_dist:

misc_info: l(3)c43. Transcripts are >= 12kb, beyond the MW marker range. Prior clone from alpha tubulin. Location is now narrowed to5E1-5 on the basis of deletion complementation: hyd is not found in

**full citation** **Mansfield, L; Biggs, J; Shearn, A. 1990 Crete meeting.**


**contact_person** **Mansfield, Liz**  **phone_number** [redacted]

**lab_address** **Biology Department**

**address_line1** **Johns Hopkins University**

**address_line2** **34th and Charles Streets**

**address_line3** **Baltimore, MD 21218**

**address_line4**


**keywords** !

DROSOPHILA GENMAPS DATABASE RECORD PRINTOUT - 8/26/92
Return corrections to John Merriam, UCLA Biology,
Los Angeles, CA 90026-1606; FAX [redacted]

source_numbe 328

full_location 85E
table_numbers 1,6 , 5 7 cross_locations

gene_name hyperplastic discs
gene_abbrev hyd
product putative >250 kD, function unknown

1 kb_cloned_genom 30 vector_genomi CosPer (Tamkun cosmid library)
kb_cloned_cDNA ~5 # vector_cDNA λgt11, λZAPII
METHOD: expression walk_jump_org YACs 0
microcloning oligonucleotide
homology x transposon_tag xxx
clonetechtext transposon tagging with P mutants --> 3 insertions; Prior clone from alpha tubulin

2 rearrangement

3 T_strain
markers

5 kb_seq_genomic ~ 5 0 acc_number_gen — kb_seq_old 0
kb_seq_cDNA ~ 3 0 acc_number_cDNA —
comments cDNA has small identity w/ PolyA+ binding protein domain of unknown function
ORF_size
ORF_functio
ORF_domain

6 kb_trans_size 12
transc_dist ~~6-7 day larvae polyA+ and total~~ all stages, greatest in embryos, pupae, adults (total RNA)

7 protein_dist all stages, greatest in emb., pup. adults

8 fusion_dist

misc_info l(3)c43. Transcripts are >= 12kb, beyond the MW marker range. Location is now narrowed to 85E1-5 on the basis of deletion complementation: hyd is not found in Df(3R)hyd-8B104, but is found in Df(3R)bY-416 i.e. hyd-8B104 does not complement, but by-416 does. We believe hyd lies nearer 85E5

full citation Mansfield, L; Biggs, J; Shearn, A. 1990 Crete meeting.


contact_person Mansfield, Liz ph [redacted] FAX [redacted]
lab_address Biology Department
address_line1 Johns Hopkins University
address_line2 34th and Charles Streets
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address_line4


keywords

a.c. MNV 12-28-92

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comments cDNA has small identity w/ Poly A+ binding protein domain of unknown function
ORF_size
ORF_functio
ORF_domain

6 kb_trans_size 12
transc_dist ~~6-7 day larvae polyA+ and total~~ all stages; greatest in embryos, pupae, adults (total RNA)

7 protein_dist ~~all stages~~; greatest in emb., pup. adults

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